

V79 COLONY FORMING ASSAY

Experiment Name : ^{210}Po toxicity (4×10^6 cell cluster, 10% labeling); Exp. # : 1;

Investigator: A. Bishayee

Date: 12/11/98

1. Set two rocker-roller at 37°C incubator with 5% CO_2 , set the Coulter Counter, wash cells (from two 150 cm^2 flusk, subcultured 1:2, 24h before) with PBS, trypsinize cells, each resuspend in 9 ml MEMB, pool, pass five times through 3 cc syringe with 21 gauge needle, perform cell count by transferring 100 ul in Coulter cup containing 20 ml isotone (Coulter balanced electrolyte solution)
2. Dilute to $\sim 400,000$ and $3,600,000$ cells/ml in MEMB [Actual count: 620,000 and 3,369,333 cells/ml]
3. Transfer 1 ml of cell suspension into ten 14 ml tubes (Falcon plastic test tube, 100×80 mm) labeled 1-10 both on cap and wall
4. Keep the tubes in the roller for 3-4 h at 37°C , 5% CO_2 Date/Time: 12/11/98; 2-00 p.m.
5. Calibrate the stock ^{210}Po -citrate
6. After 3-4 h, remove first set of test tubes ($400,000$ cells/ml) from roller and add according to

Table below.

Date/Time: 12/11/98; 4-00 p.m.

Tube #	^{210}Po -citrate uCi/ml	Cells in MEMB (ml)	MEMB (ul)	Po-citrate (12 uCi/ml) (ul)
1	0	1.0	1000	0
2	0	1.0	1000	0
3	0.1	1.0	985	15
4	0.2	1.0	965	35
5	0.3	1.0	950	50
6	0.5	1.0	915	85
7	0.8	1.0	865	135
8	1.0	1.0	835	165
9	1.2	1.0	800	200
10	1.5	1.0	750	250

7. Return test tubes to roller for 30 min.

Date/Time: 12/11/98; 4-15 p.m.

8. After 30 min, centrifuge tubes for 10 min at 2000 rpm, 4°C **Date/Time:** 12/11/98; 4:45 p.m.
9. Collect 150 ul supernatant in separate tubes
10. Resuspend in 10 ml wash MEMA
11. Centrifuge tubes for 10 min at 2000 rpm, 4°C
12. Decant supernatant, click tubes, vortex, resuspend in 10 ml wash MEMA
13. Centrifuge tubes for 10 min at 2000 rpm, 4°C
14. Decant supernatant, click tubes, vortex, resuspend in 10 ml wash MEMA
15. Centrifuge tubes for 10 min at 2000 rpm, 4°C
16. Decant supernatant, click tubes, vortex
17. Follow steps 11-16 for second set of tubes containing 3,600,000 cells, suspend in 7 ml of MEMA and transfer cells to the corresponding tubes containing 400,000 cells in step 16
18. Centrifuge tubes for 10 min at 2000 rpm, 4°C
19. Decant supernatant, click tubes, vortex, transfer the cell suspension in polypropylene microcentrifuge tubes with attached caps (Helena Plastics, 400 ul) using 200 ul pipet tips
20. Again add 200 ul ice cold MEMA, resuspend and transfer the cell suspensions in the same polypropylene microcentrifuge tubes (Total volume ~400 ul)
21. Centrifuge tubes for 5 min at 1000 rpm, 4°C
22. Transfer tubes (onto a perforated Renin pipet box) at 10°C for 72 h. **Date/Time:** 12/11/98;
23. Transfer 30 ul supernatant in three sets of vials containing 6ml liquid scintillation cocktail (Aquasol) from 150 ul supernatant removed earlier (Step 9) and count them for radioactivity **6-00 p.m.**
- Date/Time:** 12/15/98; 12:00 noon
24. After 72 h, carefully remove the supernatant from the top, resuspend pellet in 200 ul wash MEMA and transfer the content to ten 12 ml tubes (Falcon plastic test tube, 17x100 mm, labeled 1-10 both on cap and wall) containing 10 ml wash MEMA by using pasteur pipet
- Date/Time:** 12/14/98; 10:00 a.m.
25. Again add 200 ul wash MEMA in microcentrifuge tubes, resuspend and transfer the cell suspensions in 12 ml tubes
26. Centrifuge the tubes for 10 min at 2000 rpm, 4°C (precooled centrifuge)
27. Labeling and preparation of dilution tubes and colony dishes
- load 60 mm petri dishes with 4 ml MEMA
 - load 40 sterile tubes with 4.5 ml MEMA and label them 1.2, 1.3, 1.4, 1.5; 2.2, 2.3, 2.4, 2.5; X.2, X.3, X.4, X.5 etc.
28. Decant supernatant, click tubes, vortex, resuspend in 10 ml wash MEMA
29. Centrifuge tubes for 10 min at 2000 rpm, 4°C
30. Decant supernatant, click tubes, vortex, resuspend in 10 ml wash MEMA
31. Centrifuge tubes for 10 min at 2000 rpm, 4°C
32. Decant supernatant, click tubes, vortex, resuspend in 2 ml wash MEMA, pass five times

through 3 cc syringe with 21 gauge needle

33. Determine cell concentration by transferring 100 μ l to Coulter cup
 34. Vortex tube, transfer 0.5 ml into dilution tube X.5, vortex tube X.5, transfer 0.5 ml into dilution tube X.4, vortex tube X.4 and transfer 0.5 ml to tube X.3, vortex tube X.3 and transfer 0.5 ml to tube X.2 and vortex. Keep tubes on ice.
-
35. Transfer 1 ml from dilution tubes into dishes labeled X.2, X.3, X.4 (in triplicate). Only X.2 should be seeded for control test tubes.
 36. Transfer 500 μ l of cell suspension (in duplicate) to 20 ml scintillation vial containing 6 ml cocktail (Aquasol)
 37. Incubate petridishes for 1 week
 38. Count vials for radioactivity Date/Time : 12/14/98; 3-00 PM.
 39. After 1 week, wash colonies 3 times with normal (1X) saline, and 2 times with methanol. Stain colonies with 0.05% crystal violet
 40. Count colonies. There must be between 25 and 250 colonies for the dish to be a valid data point.

12/11/98

$$\begin{aligned}\text{Initial Cell Count} &= 1401, 1417, 1447 \\ \text{Avg. Cell count} &= 1421 \\ \text{Cell conc.} &= 1421 \times 4000 \\ &= 5,686,666 \text{ cells/ml}\end{aligned}$$

Dilution A : 400,000 cells/ml ; 11 ml.

$$\text{Vol. required} = \frac{4400000}{5,686,666} = 0.77 \text{ ml}$$

Take 0.8 ml cells + 10.2 ml MEMB = 11 ml.

Final count = 164, 151, 150 ; Cell conc. 620,000 cells/ml

Dilution B : 3,600,000 cells/ml ; 11 ml

$$\text{Vol. required} = \frac{39600000}{5686666} = 6.9 \text{ ml}$$

Take 7ml cells + 4ml MEMB = 11 ml.

Final count = 852, 853, 822 ; Cell conc. = 3,369,333 cells/ml

1M Hcl : 1M Citrate

1ml Hcl :

100µl Hcl : 100µl Citrate

: 200 "

500 "

1ml "

$$3 \text{ ml} \times 2 \text{ M} = 6 \text{ M} \times \text{X}$$

300µl
1.5 ml

$$3 \text{ ml} \times 6 \text{ M} =$$

1.8 ml

Stock 100µCi/ml

1:5 \Rightarrow 16.6µCi/ml

2M Hcl : 1M Na-Citrate

1:1

1:2

1:3

1:4

1:5

1:9

pH

3

4

5

6

6.8 ✓

7

4µCi

1:5

100µCi - 6M

16µCi/ml

1:5

① change ratio 2

② ~~to~~ higher survival 2

③

²¹⁰Po- tartrate, 10% labeling

Expt #1

Stock on 10/06/98 = 100 μ ci/ml.

12/11/98

$$A_t = A_0 \times e^{-\lambda t}$$

$$= 100 \times e^{-\frac{0.693 \times 65}{138}}$$

$$= 100 \times 0.72$$

$$= 72 \mu\text{ci/ml. on } 12/11/98$$

After 1:5 dilution: ²¹⁰Po-citrate = 12 μ ci/ml.

For this: Take 1.5 ml 1M Na-citrate in 1.8 ml Nalgene Orvovial
Add 300 μ l of stock Po-cit in 2M Hcl

X 3

	μ ci/ml	Cells	MEMB (ml)	Po-citrate (12 μ ci/ml)
1	0	20	1000	0
2	0	20	1000	0
3	0.1	10	0.984	0.016
4	0.2	20	0.967	0.02 0.033
5	0.3	30	0.950	0.05
6	0.5	20	0.917	0.083
7	0.8	20	0.867	0.133
8	1.0	20	0.834	0.166
9	6.0 1.2	20	0.8	0.2
10	1.5	20	0.75	0.25

USER: 5 ID:PO-210 PRESET TIME: 1.00 TUE 15 DEC 1998 12:19
 SAMPLE REPEAT: 1 CYCLE REPEAT: 1 SCR:N R6232:N
 H#: 1 ADC:N QDF:N RCH:N
 CHANNEL 1-LL:600 UL: 900 ZSIGMA: 2.00 BKG SUB: 0.00 BKG ZSIG: 0.00 LSR:
 DATA CALC: CPM, UNKNOWN REPLICATES: 1 NORM FACTOR: 0 1.00000
 HALF LIFE(DAYS):N

No Potentials (10% label)

Expt # 1

20ml medium

SAM	POS	CH	CPM	ZSIG%	TIME	EL TIME	AVG	H#	ERR
1	**	1	10.00	63.25	1.00	1.60	55.0		
2	**	2	2.00	141.4	1.00	3.28	56.0		
3	**	3	12.00	57.74	1.00	4.97	55.0		
4	**	4	12.00	57.74	1.00	6.65	55.0		
5	**	5	1916.00	4.57	1.00	8.33	54.0		
6	**	6	1808.00	4.70	1.00	10.01	55.0		
7	**	7	1918.00	4.57	1.00	11.64	61.0		
8	**	8	1801.00	4.71	1.00	13.33	59.0		
9	**	9	3424.00	3.42	1.00	15.00	56.0		
10	**	10	3809.00	3.24	1.00	16.68	56.0		
11	**	11	5166.00	2.78	1.00	18.32	58.0		
12	**	12	5912.00	2.60	1.00	20.00	56.0		
13	**	1	14184.00	1.94	0.75	21.49	57.0		
14	**	2	26789.33*	1.92	0.38	22.58	54.0		
15	**	3	14722.76	1.94	0.72	24.03	56.0		
16	**	4	16667.69	1.92	0.65	25.36	54.0		
17	**	5	53644.44	1.82	0.23	26.30	53.0		
18	**	6	66993.33	2.00	0.15	27.12	56.0		
19	**	7	12517.58	1.97	0.62	28.67	55.0		
20	**	8	14182.67	1.94	0.75	30.10	55.0		
21	**	9	91206.66	1.71	0.15	30.81	53.0		
22	**	10	106085.72	1.47	0.17	31.71	54.0		

10M

1

3M-G: 20ml glass scintillation vial + 6ml Aquasol
3M-P: 20ml plastic " " + " "

** possibly radioactivity added twice*

Ⓚ error in addition
†† error in addition

TABLE-1

Expt. #: ~~12/15/98~~ 1

Date/Time: 12/15/98; 12-20 p.m.

Tube #	Medium count for <u>90</u> ul (cpm)	Avg. cpm	dpm [cpm/1]	μ Ci/ml (A_c) on counting [dpm/ 66600 ⁴⁴⁴⁰⁰]	μ Ci/ml (A_o) on addition [$A_c/e^{-\lambda t}$]
1	See the				
2	attached sheet				
3		1859.5	1859.5	0.0418	
4		3616.5	3616.5	0.0814	
5		5539	5539	0.1247	
6		14324	14324	0.3226	
7		15694	15694	0.3534	
8		60318	60318	1.35	
9		13349	13349	0.300*	
10		98645	98645	2.22	

* error in addition

210Po toxicity (10% labeling)

Expt #01

F-451 / Rao/Honell

500µl cells

PAGE: 1

USER: S ID:PO-210 PRESET TIME: 1.00 MON 14 DEC 1998 14:55
SAMPLE REPEAT: 1 CYCLE REPEAT: 1 SCR:N RS232:N
R#: 1 AGC:N DCF:N RCM:N
CHANNEL: 1-LL:500 UL: 900 2SIGMA: 2.00 BKG SUB: 0.00 BKG 2SIG: 0.00 LSR: 0
DATA CALC: CPM, UNKNOWN REPLICATES: 1 NORM FACTOR:Q 1.00000
HALF LIFE(DAYS):N

SAM	POS	CH	CPM	2SIG%	TIME	EL TIME	AVG H#	ERR
1	**	1	1c 6.00	81.65	1.00	1.55	105.0	
2	**	2	2c 17.00	48.51	1.00	3.27	108.0	
3	**	3	3c 214.00	13.67	1.00	4.91	106.0	
4	**	4	3c 201.00	14.11	1.00	6.63	106.0	
5	**	5	4c 173.00	15.21	1.00	8.37	109.0	
6	**	6	4c 152.00	16.22	1.00	10.04	107.0	
7	**	7	5c 208.00	13.87	1.00	11.72	106.0	
8	**	8	5c 230.00	13.19	1.00	13.35	106.0	
9	**	9	6c 309.00	11.38	1.00	15.03	106.0	
10	**	10	6c 327.00	11.03	1.00	16.67	102.0	
11	**	11	7c 397.00	10.04	1.00	18.40	98.0	
12	**	12	7c 403.00	9.96	1.00	20.08	97.0	
13	**	1	8c 684.00	7.65	1.00	21.87	102.0	
14	**	2	8c 711.00	7.50	1.00	23.61	98.0	
15	**	3	ac 371.00	10.38	1.00	25.24	98.0	
16	**	4	ac 396.00	10.05	1.00	26.87	100.0	
17	**	5	10c 653.00	7.83	1.00	28.56	99.0	
18	**	6	10c 607.00	8.17	1.00	30.24	99.0	

TABLE-2

Expt. # : j

Date/Time : 12/14/98; 3-00 p.m.

Tube #	Radioactivity for 500 ul cell suspension (cpm)	Avg. cpm	dpm [cpm/1]	μ Ci/ml (A) on counting [dpm/111x10 ⁴]	μ Ci/ml (A ₀) after 12 h incubation [A/e ^{-λt}]
1	<i>See the attached sheet</i>				
2					
3		207.5	207.5	0.000186	
4		162.5	162.5	0.000146	
5		219	219	0.000197	
6		319	319	0.000287	
7		400	400	0.000360	
8		698	698	0.000628	
9		383.5	383.5	0.000345	
10		630	630	0.000567	

TABLE-3

Expt. # : 1

Date/Time : 12/14/98

Tube #	Coulter count for 100 ul cell suspension	Avg. count	Cells/ml [Avg. count x 400 10]	fCi/cell [uCi/ml x 10 ⁹ Cells/ml]
1	543, 516, 541	533	2133333	-
2	500, 502, 513	505	2020000	-
3	460, 481, 434	458	183333	0.1014
4	531, 542, 504	525	2102666	0.0694
5	531, 493, 488	504	2016000	0.0977
6	542, 530, 518	530	2120000	0.1353
7	512, 473, 457	480	1922666	0.1872
8	514, 499, 487	500	2000000	0.314
9	454, 439, 444	445	1782666	0.1935
10	469, 490, 450	469	1878666	0.3018

TABLE-4

Expt #: 1

Date: 12/21/98

Colony Counts and Survival Fraction

Tube.dilution	Colony 1	Colony 2	Colony 3	Avg Colony	SF
1.2	156	166	142	} 146.83	—
2.2	139	149	129		—
3.2	137	130	123	130	0.8849
4.2	132	140	151	141	0.9598
5.2	121	125	129	125	0.8509
6.2	108	100	117	100.33	0.7374
7.2	83	91	99	91	0.6194
8.2	56	64	52	57.33	0.3962
9.2	76	70	82	76	0.5173
10.	79	72	65	72	0.4901