## **V79 COLONY FORMING ASSAY**

8×105 cells in 2 ml, acute

Experiment Name: 137Cs toxicity; Exp. #:1; Investigator: A. Bishayee; Date: 09/07/98

- Set the rocker-roller at 37°C incubator with 5% CO<sub>2</sub>, set the Coulter Counter, wash cells (from one 75 cm² flusk, subcultured 1:2, 24h before) with PBS, trypsinize cells, resuspend in 6 ml MEMB for each flusk, pool, vortex, pass five times through 3 cc syringe with 21 gauge needle, perform cell count by transfering 100 ul in Coulter cup containing 20 ml isotone (Coulter balanced electrolyte solution)
- 2. Dilute to ~400,000 cells/ml in MEMB (final volume 11 ml) [Actual count: \(\frac{369}{17\times 000}\) cells/ml)
- 3. Transfer 1 ml of cell suspension into ten 12 ml tubes (Falcon plastic test tube, 12x175 mm) labeled 1-10 both on cap and wall
- 4. Roll the tubes for 15 h at 37°C, 5% CO<sub>2</sub> Date/Time: 09/07/98; 6-00 p.m.
- 5. After ~15 h incubation period, remove tubes, add 10 ml wash MEMA, vortex and centrifuge at 2000 rpm at 4°C for 10 min (precooled centrifuge). Date/Time: 09/08/98; 10-00 a.m.
- 6. Decant supernatant, click tubes, vortex, resuspend in 10 ml wash MEMA
- 7. Centrifuge tubes for 10 min at 2000 rpm, 4°C
- 8. Decant supernatant, click tubes, resuspend in 10 ml wash MEMA
- 9. Centrifuge tubes for 10 min at 2000 rpm, 4°C
- 10. Deacnt supernatant, click tubes, resuspend in 2 ml cold MEMA
- 11. Transfer tubes at 10°C for 72 h. Date/Time: 09/08/98; 11-00 a.m.
- 12. After 72 h, place the tubes on the perforated box containing ice (to maintain ~ 10.5°C)
- 13. The tubes were irradiated using Mark I irradiator (137Cs gamma-ray), one tube at a time, while placing onto a box containing ice as per the Table below

Tube#	Total Dose (Rad)	Dose rate (Rad/min)	Time (min)	Attenuat.
1	0	0	0	0
2	0	0	0	0
3	200	165	1.21	X-5
4	400	165	2.42	X-5
5	600	165	3.64	X-5
6	800 -	389	2.06	X-2
7	1000 -	389	2.57	X-2
8	1200 ·	749.6	1.60	0
9	1600 ·	749.6	2.13	0
10	2000 -	749.6	2.67	0

- 14. After irradiation, add 8 ml wash MEMA in each tube
- 15. Centrifuge the tubes for 10 min at 2000 rpm, 4°C (precooled centrifuge)
- 16. Labeling and preparation of dilution tubes and colony dishes
  - -load 57 60 mm petri dishes with 4 ml MEMA
  - load 39 T-tubes with 4.5 ml MEMA and label them 1.2, 1.3, 1.4, 1.5; 2.2, 2.3, 2.4, 2.5; X.2, X.3, X.4, X.5 etc.
  - 17. Decant supernatant, click tubes, vortex, resuspend in 10 ml wash MEMA
- 18. Centrifuge tubes for 10 min at 2000 rpm, 4°C
- 19. Decant supernatant, click tubes, vortex, resuspend in 2 ml wash MEMA, pass five times through 3 cc syringe with 21 gauge needle
- 20. Determine cell concentration by transfering 100 µl to Coulter cup
- 21. Vortex tube, transfer 0.5 ml into dilution tube X.4, vortex tube X.4 and transfer 0.5 ml to tube X.3 and vortex tube X.3 and transfer 0.5 ml to tube X.2. Keep tubes on ice.
- 22. Transfer 1 ml from dilution tubes into dishes labeled X.2, X.3, X.4 (in triplicate). Only X.2 should be seeded for control T-tubes.
- 23. Incubate petridishes for 1 week
- 24. After 1 week, wash colonies 3 times with normal (1X) saline, and 2 times with methanol. Stain colonies with 0.0 5% crystal violet
- 25. Count colonies. There must be between 25 and 250 colonies for the flask to be a valid data point.

9/7/98

Avg. all count = 1874, 1991, 1946 Avg. all count = 1937 cell come. = 1937 x 400 = 774800 cell/re/

For dilusion,

vol. of cell suppersion required =  $\frac{4400000}{774800}$  we =  $\frac{4400000}{2}$  =  $\frac{4400000}{5.67}$  me

Take 5.7 ml cellnipumon + 5.3 ml HEMB = C/nl

Aftér dilusion,

Final Count = 965, 980, 975 Arg. Count = 973 Cell Core = 389, 333 Cells/ml

TABLE-4

Expt. #: 1

Date: 09/19/98

## Colony Counts and Survival Fraction

Tube.dilution	Colony 1	Colony 2	Colony 3	Avg Colony	SF
1.2	110	119	125	(120.33	
2.2	120	132_	116	<u> </u>	
3.2	105	96	87	96	0. 7977
4.2	70	54	63	62. 33	0.5180
5.2	40	28	35	34.33	0.2853
6.3	48	52	44	4.8	0.0398
7.4	58	39	49	0.48	0.004
8.4	15	13	16	0.1466	0.0012
9.4	2	3	9	0.03	0-00024
10.4	1	2_	1	0.01	0.00011

