V79 COLONY FORMING ASSAY

Experiment Name : Cell separation by FACS and SF (^{3}HTdR cluster, 50% labeling, three ^{3}HTdR conc.) **Exp. # :** 1; **Investigator:** A. Bishayee **Date:** 09/09/99

1. Set the rocker-roller at 37^{0}C incubator with 5% CO_2, set the Coulter Counter, wash cells (from two 150 cm^2 flusk, subcultured 1:2, 24h before) with PBS, trypsinize cells, each resuspend in 9 ml MEMB, pool, pass five times through 3 cc syringe with 21 gauge needle, perform cell count by transfering 100 ul in Coulter cup containing 20 ml isotone (Coulter balanced electrolyte solution) OR 6,000,000
2. Dilute to ~3,000,000 cells/ml in MEMB [Actual count : cells/ml)
3. Transfer 1 ml of cell suspension into 12 ml tubes (Falcon plastic test tube, 17x100 mm) labeled 1-8 both on cap and wall
4. Keep the tubes in the roller for 3-4 h at 37°C, 5% CO_2 **Date/Time:** 09/09/99 ; 3-00 pm
5. Prepare MEMB containing radioactivity in hood

   60 µl ^{3}HTdR (Stock : 1 µCi/µl on 6/17/99) + ml MEMB
6. After 3-4 h, remove tubes from roller and add MEMB with or without radioactivity according to Table below. **Date/Time:** 09/09/99, 7-15 pm.

Tube # Pl. See attached sheet for details	^{3}HTdR uCi/ml	Cells in MEMB (ml)	MEMB (ml)	MEMB+ ^{3}HTdR (ml) [20uCi/ml]	CFDA in PBS (0.05 uM) (ml)	Treatment * See details
1	0	1.0	1.0	0	0	None
2	0	1.0	1.0	0	2	100% dyed
3	0	1.0	1.0	0	2	50% dyed
4	0	1.0	1.0	0	2	50% dyed
5	1	1.0	0.9	0.1	2	50% dyed, labeled
6	1	1.0	0.9	0.1	2	50% " , "
7	3	1.0	0.7	0.3	2	50% dyed, "
8	3	1.0	0.7	0.3	2	50% dyed, "
9	6	1.0	0.4	0.6	2	50% dyed "
10	6	1.0	0.4	0.6	2	50% dyed "

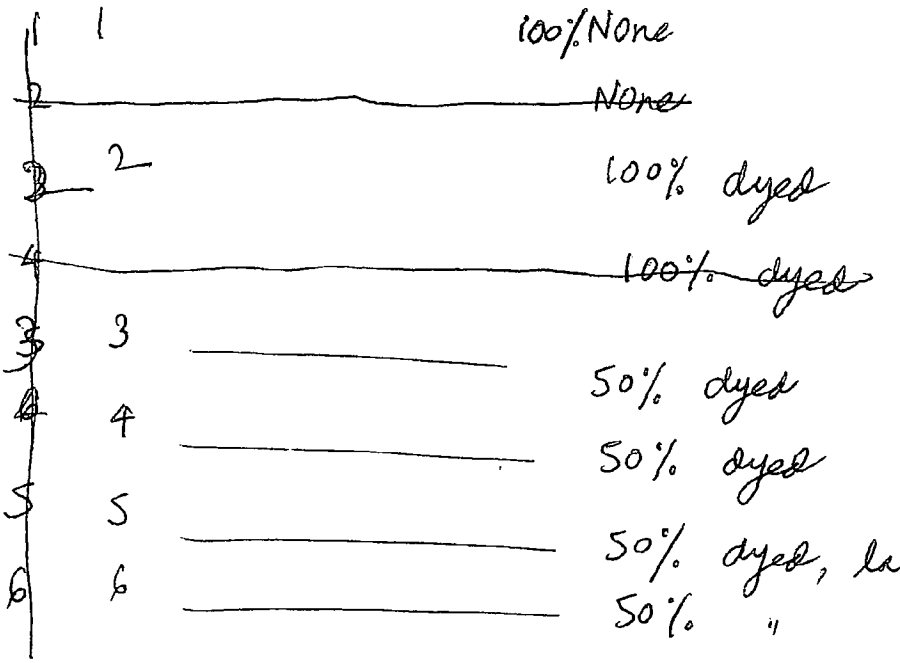
7. Return test tubes to roller for 12 h . **Date/Time:** 09/09/99; 7-30 p.m. ~~09/10/99; 9-30 a.m.~~
8. Next day, while test tubes are in roller label 10 gamma-tubes (13 X 100 mm VWR glass test tube)
9. After ~12 h incubation period, remove tubes and centrifuge at 2000 rpm at 4°C for 10 min (***precooled centrifuge***) . **Date/Time:** ~~09~~ 09/10/99; 9-00 a.m.
10. Remove buckets from centrifuge and carefully remove 150 µl of supernatant and place in prelabeled gamma-tube.
11. Decant supernatant, click tubes, vortex, resuspend in 10 ml wash MEMA
12. Centrifuge tubes for 10 min at 2000 rpm, 4°C
13. Decant supernatant, click tubes, vortex, resuspend in 10 ml wash MEMA
14. Centrifuge tubes for 10 min at 2000 rpm, 4°C
15. Decant supernatant, click tubes, vortex, resuspend in 10 ml wash MEMA
16. Centrifuge tubes for 10 min at 2000 rpm, 4°C
17. Decant supernatant, click tubes, vortex, resuspend in ~~2 ml of PBS, syringe and perform cell count as well as radioactivity count by transferring aliquots~~.
18. ~~Add~~ 8 ml of PBS in each tube, vortex and transfer the content to 15-ml plastic centrifuge tube
18. Centrifuge tubes for 10 min at 2000 rpm, 4°C
19. Decant supernatant, click tubes,vortex
20. Add 2 ml of 0.05 uM CFDA in prewarmed PBS as per the Table and PBS in the remaining tubes.
21. Incubate all tubes at 37°C for 15 min.
21. Centrifug tubes for 10 min at 2000 rpm, 4°C
22. Decant supernatant, click tubes, vortex, add 2 ml prewarmed MEMA
23. Incubate all tubes at 37°C for 30 min.
24. Centrifuge and decant the supernatant, suspend in 5 ml MEMA
25. Transfer the content of one tube to the corresponding tube
26. Centrifuge, decant the supernatant
27. Transfer the cell suspension in polypropylene microcentrifuge tubes with attached caps (Helena Plastics, 400 ul) using 200 ul pipet tips
28. Again add 200 ul ice cold MEMA, resuspend and transfer the cell suspensions in the same polypropylene microcentrifuge tubes (Total volume ~400 ul)
29. Centrifuge tubes for 5 min at 1000 rpm, 4°C
30. Transfer tubes at 10°C for 72 h. **Date/Time:** 11-00 a.m/ 09/10/99
31. After 72 h, carefully remove the supernatant from the top, resuspend pellet in 200 ul wash MEMA and transfer the content to ~~eight~~ 10 15 ml tubes containing 10 ml PBS by using pasteur pipet **Date/Time:** 09/10/99; 1-30 p.m.

32. Again add 200 ul PBS in microcentrifuge tubes, resuspend and transfer the cell suspensions in 15 ml tubes
33. Centrifuge the tubes for 10 min at 2000 rpm, 4°C ***(precooled centrifuge)***
32. Decant supernatant, click tubes, vortex, pooled cells from corresponding tubes, centrifuge, decant the suoernatant, resuspend in 2 ml PBS with 0.005 mM EDTA, syringe and transfer aliquots (100 ul) for cell count and radioactivity count
33. Centrifuge, decant, resuspend in 1 ml PBS with 0.005 mM EDTA to have ~10,000,000 cells/ml for each tube and transfer ~1ml in Falcon 12x75 mm polystyrene 6 ml tube, wrap the tubes with aluminium foil, put in ice and transfer for FACS study.

Tube #p

(cluster of ~~6~~ million cells)

Treatment



1 1 — 100% None

~~2 — None~~

2 — 100% dyed

~~4 — 100% dyed~~

3 — 50% dyed

4 — 50% dyed

5 — 50% dyed, labeled with ^3H (conc. 1) 1 μCi/ml

6 — 50% " " " " "

7 — 50% dyed, labeled with ^3H (conc. 2) 3 μCi/ml

8 — 50% " " " " "

9 — 50% dyed, labeled with ^3H (conc. 3) 6 μCi/ml

10 — 50% dyed, labeled with ^3H (conc. 3)

Tube #	Treatment
1	None
2	100% dyed
3	50% dyed, [illegible]
4	50% dyed
5	50% dyed

Tube #	# of cells (million)	CFDA
1	6	0
2	6	2
3	3	2
4	3	0
5	3	2
6	3	0
7	3	2
8	3	0
9	3	2
10	3	0
11	3	2
12	3	0
13	3	2
14	3	0
15	3	2
16	3	0
17	3	2
18	3	0

Take 60μl 3H + 3ml MEMB = 20μci/ml

	MEMB	MEMB + 3H (20μci/ml)
5A 6A	0.9	0.7
7A 8A	0.4	0.8
9A 10A	6μl	0.4 0.6

2 μl

200μci = 1ml
2 — 1/10

Prepare 20ml 0.05 μM LPDA in 1X PBS

30 μl medium

[illegible] 5 ID-H3 HOWELL   PRESET TIME: 1.00   FRI 10 SEP 1999 13:42
SAMPLE REPEAT: 1 CYCLE REPEAT: 1 COR.N   RS232:N
[illegible] 1 PBD:N [illegible] ROM:N
[illegible] 2S%: 2.00 AKG SUB: 0.00 BKG SUB: [illegible]
DATA CALC: DPM, UNKNOWN REPLICATES: 1   NORM FACTOR:G 1.00000
[illegible] LIFE(DAYS):N

SMPL	POS	CH		CPM	2S%	TIME	EL TIME	AVG H#	ERR
1	##-1	1		20433.33	1.93	0.62	1.47	82.0	
2	##-2	1	5M	1[illegible].60	1.95	0.62	3.37	77.0	
3	##-3	1	6M	15337.14	1.93	0.70	5.25	81.0	
4	##-4	1		15600.00	1.96	0.67	6.73	80.0	
5	##-5	1	7M	36551.1[illegible]	1.77	0.23	8.23	82.0	
6	##-6	1		*7.00	73.39	1.00	10.30	80.0	
7	##-7	1	8M	42175.00	1.99	0.24	11.66	83.0	
8	##-8	1		[illegible]2401.37	1.39	0.2[illegible]	12.94	85.0	
[illegible]			9M	[illegible]102.56	1.65	0.17	14.19	82.0	
[illegible]		1		[illegible]66.22	1.76	0.14	15.[illegible]	83.0	
11	##-11	1	10M	[illegible]6515.96	1.4[illegible]		17.[illegible]	83.0	
12	##-12	1		36626.66	1.73	0.15	18.60	23.0	
13	##-13						19.03		101
14	##-14						19.45		10[illegible]

* Sample was not added by mistake!

B011680

Cells suspended in 2 ml 09/11/99 : 50 µl

1.	712, 711, 702	2·8 M/ml	Take 10µl for radio activity count
2	808, 795, ~~789~~, 741, 753, 720	2·9 M/ml	
3	1503, 1524, 1511	6·0 M/ml	
4	1547, 1516, 1539	6·1 M/ml	
5	1509, 1556, 1519	6·1 M/ml	
6	1401, 1426, 1435	5·7 M/ml	

45, 866 cells/ml

After sorting

Cells suspended in 1 ml : 500 µl MS² MEMA

1.	110, 113, 119	45, 866 cells/ml	1·3 ml
2	122, 107, 112	45, 466 cells/ml	1·2 ml
3	141, 145, 135	54, 533 cells/ml	1·8 ml
4	95, 105, 111	41, 466 cells/ml	1·2 ml

Take 0·1 ml for radioactivity count

i) Dilutions were made to have 20,000 cells/ml
ii) Take 0·5 ml + 4·5 ml MEMA = 5 ml (2,000 cells/ml)
iii) Take 0·5 ml from ii + 4·5 ml MEMA = 5 ml (200 cells/ml)

iv) Plate ~~200 cells/ml~~ 1 ml (200 cells) for Conc 1 X3
v) Plate 1 ml (200 or 2000) for Conc 2-4 X3

Initial count = 1010, 1020,

Avg count =

Cell conc =

22 ml of 3,000,000 cells/ml = 66,000,000 cells

Vol. required =

Take ml cells + ml MEMB = 22 ml

Avg count =

Cell conc =

B011682

TABLE-1

Expt. # : 1 **Date/Time :** 09/10/99; 1-45 p.m.

Tube #	Medium count for 30 ul (cpm)	Avg. cpm	dpm [cpm/0.65]	μ Ci/ml (A_t) on counting [dpm/66600]	μ Ci/ml (A_o) on addition [$A_t/e^{-\lambda t}$]
1					
2					
3					
4					
5		18407	28319	0·425	
6		15488	23797	0·357	
7		56551	87001	1·31	
8		42291	65063	0·98	
9		89123	137113	2·06	
10		87770	135031	2·03	

Input header cells

USER: A ID:H3 HOWELL PRESET TIME: 1.00 TUE 14 SEP 1999 11:28
SAMPLE REPEAT: 1 CYCLE REPEAT: 1 SCR:N RS232:N
H# AQC:N DCF:N RCM:N
CHANNEL 1-LL: 0 UL: 400 2SIGMA: 2.00 BK3 SUB: 0.00 BK3 2SIG: 0.00 LSR: 0
DATA CALC: CPM, UNKNOWN REPLICATES: 1 NORM FACTOR:0 1.00000
HALF LIFE(DAYS):N

SAM	POS	CH		CPM	2SIG%	TIME	EL TIME	AVG H#	ERR
1	**-1	1		12.00	57.74	1.00	1.89	81.0	
2	**-2	1	3,4c	8.00	70.71	1.00	3.96	81.0	
3	**-3	1		888.00	6.71	1.00	6.08	82.0	
4	**-4	1	5,6c	923.00	6.38	1.00	8.32	81.0	
5	**-5	1		2657.00	3.68	1.00	10.55	82.0	
6	**-6	1	7,8c	2669.00	3.67	1.00	12.73	81.0	
7	**-7	1		9759.00	2.64	1.00	14.96	82.0	
8	**-8	1	9,10c	9426.00	2.72	1.00	16.98	80.0	
9	**-9	1		123.00	18.03	1.00	19.05	84.0	
10	**-10	1		126.00	17.82	1.00	21.02	82.0	
11	**-11	1	*	78.00	22.65	1.00	23.03	84.0	
12	**-12	1		83.00	21.95	1.00	25.06	84.0	
13	**-13	1		22.00	42.64	1.00	27.03	82.0	
14	**-14	1		14.00	53.45	1.00	29.01	84.0	
15	**-15	1		10.00	63.25	1.00	31.19	83.0	
16	**-16	1		11.00	60.30	1.00	33.42	84.0	

~~100~~ 10 µl cells before cell sorting (50% labelled, dyed cells)

100 µl cells following sorting (unlabeled + undyed cells)

* S. Virds were touched with gloves.

$$\mu ci/ml = \frac{905}{0.65 \times 60 \times 31000}$$

$$=$$

TABLE-2

Expt. # : 1 **Date/Time :** 09/14/99; 11-30 a.m.

Tube #	Radioactivity for ~~200~~ 100 ul cell suspension (cpm)	Avg. cpm	dpm [cpm/0.65]	µ Ci/ml (A_t) on counting [dpm/~~444000~~ 222000]	µ Ci/ml (A_o) after 12 h incubation [$A_t/e^{-\lambda t}$]
1					
2					
3 }	0	~~985~~ 0			
4 }					
5 }		9050	1392		0.0063
6 }					
7 }		26630	2663		0.012
8 }					
9 }		55920	8603		0.039
10 }					

TABLE-3

Expt. # : **Date/Time :**

Tube #	Coulter count for 100 ul cell suspension	Avg. count	Cells/ml [Avg. count x 4000]	pCi/cell [uCi/ml x10^6 Cells/ml]	nCi/cluster [pCi/cell x 4000]
1	712, 711, 702	708	2,833,333	–	
2	741, 753, 720	738	2,952,000	–	
3	1503, 1524, 1511	1512	6,050,666		
4					
5	1547, 1518, 1539	1534	6138666	0.0010	4
6					
7	1509, 1556, 1519	1528	6,112,000	0.0019	7.6
8					
9	1401, 1426, 1435	1420	5,682,666	0.0069	27.6
10					

3

4

5

6

	mRad/cell (50% label)	mRad/cc	KRad/cluster [nCi/cc x 0.037]
4	0.37	0.74	0.148
5	0.7	1.4	0.281
6	2.5	5.0	1.02

TABLE-4

Expt # : 1 **Date :** 09/20/99

Tube.dilution	Colony 1	Colony 2	Colony 3	Avg Colony for 4.2	SF
3.2	85	72	92	83	
4.2	15	21	12	16	
5.2	7	5	6	6	
6.3	3	1	4	0.26	

Sorted cells

TABLE-2

Expt. # : 1 **Date/Time :** 05/25/00

Tube #	Radioactivity for ~~200~~ 100 ul cell suspension (cpm)	Avg. cpm	dpm [cpm/0.65]	µCi/ml (A_t) on counting [dpm/~~444000~~ 222000]	µCi/ml (A_o) after 12 h incubation [$A_t/e^{-\lambda t}$]
1					
2					
3	22, 14	18	27.7	0.00012	
4	11, 10	10.5	16.2	0.00007	
5					
6					
7					
8					
9					
10					

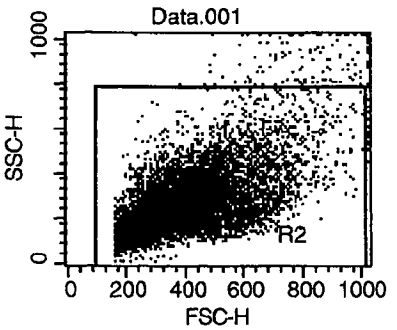
Sorted cells

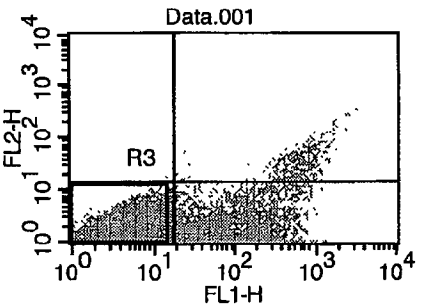
TABLE-3

Expt. # : 1 **Date/Time :** 05/25/00

Tube #	Coulter count for 100 ul cell suspension	Avg. count	Cells/ml [Avg. count x 4000]	pCi/cell [uCi/ml x10^6 Cells/ml]	mBq/cell
1					
2					
3	129, 145, 135	136	54, 533	0.0022	0.06
4	95, 105, 114	103.6	41, 466	0.0017	0.06
5					
6					
7					
8					
9					
10					

Ampicin





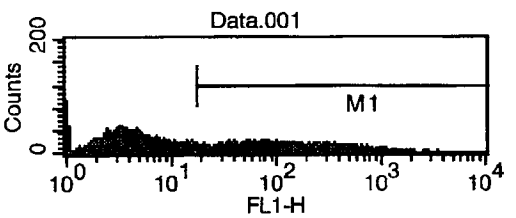
Quadrant Statistics

File: Data.001
Sample ID:
Tube:
Acquisition Date: 13-Sep-99
Gated Events: 9783
X Parameter: FL1-H (Log)
Quad Location: 18, 14

Log Data Units: Linear Values
Patient ID:
Panel:
Gate: G2
Total Events: 10000
Y Parameter: FL2-H (Log)

unstained

Quad	Events	% Gated	% Total	X Mean	X Geo Mean	Y Mean	Y Geo Mean
UL	1	0.01	0.01	14.33	14.33	18.27	18.27
UR	357	3.65	3.57	734.63	621.18	43.41	34.63
LL	5739	58.66	57.39	5.19	4.17	1.95	1.67
LR	3686	37.68	36.86	157.36	99.92	2.24	1.66



Histogram Statistics

File: Data.001
Sample ID:
Tube:
Acquisition Date: 13-Sep-99
Gated Events: 9783
X Parameter: FL1-H (Log)

Log Data Units: Linear Values
Patient ID:
Panel:
Gate: G2
Total Events: 10000

Marker	Left, Right	Events	% Gated	% Total	Mean	Geo Mean	CV	Median	Peak Ch
All	1, 9910	9783	100.00	97.83	89.14	16.57	215.33	8.98	1
M1	17, 9910	4099	41.90	40.99	205.73	114.43	123.48	103.66	52

09/13/99

① Threshold

+1281677 – Total

633918 sorted #1

② Threshold Total +

306115 sorted

295463 #2

③

2832454 Threshold Total

616260 – sorted #3

④

116,800? sorted

300152

416152 #4

1000 ml 1M = 529.29 g

1000 ml 1mM = 0.53 g = 530 mg

1 ml 1mM = 0.530 mg = 530 µg

~~[illegible]~~ 1 ml 10mM = 5.3 mg

~~5 ml — 2.6 mg~~

1000 ml 1M = 960 g

1000 ml 1mM = 0.96 g

~~1000 ml 10 mM = 9.6 g~~

10 ml 10 mM = 0.096 g

= 96.9 mg

1 ml 10mM = 9.7 mg

$S_1V_1 = S_2V_2$

~~10~~ µM × V_1 = 10,000 × 10

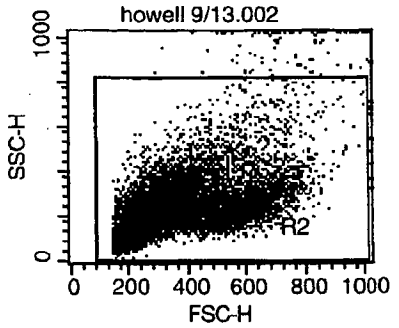
10,000 × V_1 = 10 × 10

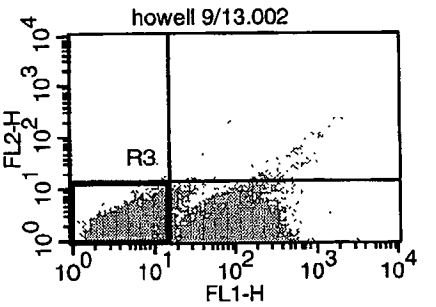
5.3 — 1

1 — 1/5.3

3-43

200,000 → 20,000 → 2,000 → 200



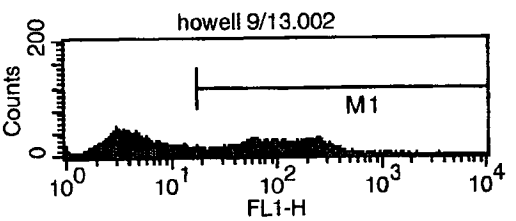


Quadrant Statistics

File: howell 9/13.002
Sample ID:
Tube:
Acquisition Date: 13-Sep-99
Gated Events: 9877
X Parameter: FL1-H (Log)
Quad Location: 16, 14

Log Data Units: Linear Values
Patient ID:
Panel:
Gate: G2
Total Events: 10000
Y Parameter: FL2-H (Log)

Quad	Events	% Gated	% Total	X Mean	X Geo Mean	Y Mean	Y Geo Mean
UL	4	0.04	0.04	13.28	13.19	21.56	20.86
UR	72	0.73	0.72	488.95	352.53	43.17	30.77
LL	4997	50.59	49.97	5.15	4.38	2.16	1.66
LR	4804	48.64	48.04	125.95	92.24	2.41	1.83

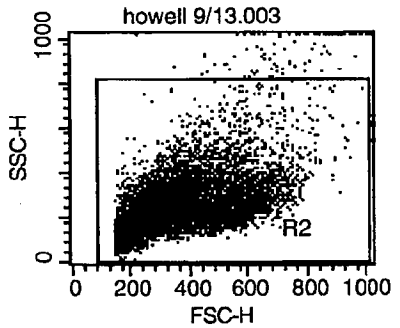


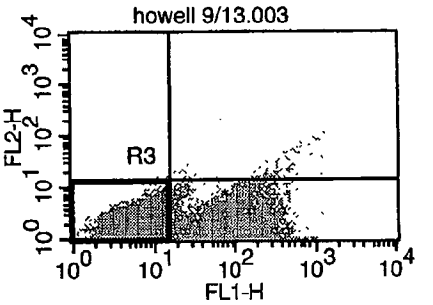
Histogram Statistics

File: howell 9/13.002
Sample ID:
Tube:
Acquisition Date: 13-Sep-99
Gated Events: 9877
X Parameter: FL1-H (Log)

Log Data Units: Linear Values
Patient ID:
Panel:
Gate: G2
Total Events: 10000

Marker	Left, Right	Events	% Gated	% Total	Mean	Geo Mean	CV	Median	Peak Ch
All	1, 9910	9877	100.00	98.77	67.43	19.92	160.98	14.46	2
M1	17, 9910	4812	48.72	48.12	132.84	96.28	94.73	97.34	55



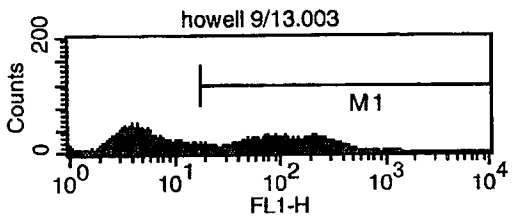


Quadrant Statistics

File: howell 9/13.003
Sample ID:
Tube:
Acquisition Date: 13-Sep-99
Gated Events: 9886
X Parameter: FL1-H (Log)
Quad Location: 16, 14

Log Data Units: Linear Values
Patient ID:
Panel:
Gate: G2
Total Events: 10000
Y Parameter: FL2-H (Log)

Quad	Events	% Gated	% Total	X Mean	X Geo Mean	Y Mean	Y Geo Mean
UL	2	0.02	0.02	12.72	12.58	16.70	16.70
UR	80	0.81	0.80	390.86	270.08	29.77	25.05
LL	4897	49.53	48.97	5.48	4.73	2.04	1.75
LR	4907	49.64	49.07	131.27	95.07	2.40	1.80

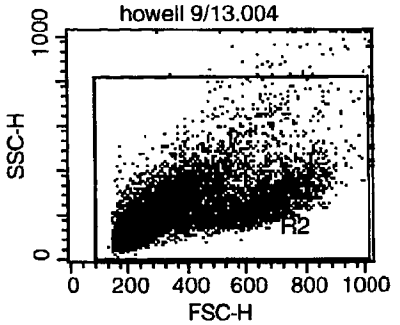


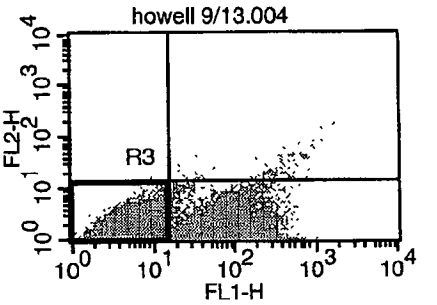
Histogram Statistics

File: howell 9/13.003
Sample ID:
Tube:
Acquisition Date: 13-Sep-99
Gated Events: 9886
X Parameter: FL1-H (Log)

Log Data Units: Linear Values
Patient ID:
Panel:
Gate: G2
Total Events: 10000

Marker	Left, Right	Events	% Gated	% Total	Mean	Geo Mean	CV	Median	Peak Ch
All	1, 9910	9886	100.00	98.86	71.03	21.68	148.95	16.85	3
M1	17, 9910	4914	49.71	49.14	137.20	99.24	85.64	99.10	70



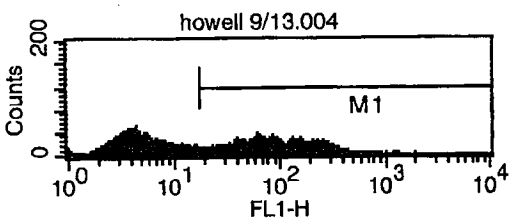


Quadrant Statistics

File: howell 9/13.004
Sample ID:
Tube:
Acquisition Date: 13-Sep-99
Gated Events: 9894
X Parameter: FL1-H (Log)
Quad Location: 16, 14

Log Data Units: Linear Values
Patient ID:
Panel:
Gate: G2
Total Events: 10000
Y Parameter: FL2-H (Log)

Quad	Events	% Gated	% Total	X Mean	X Geo Mean	Y Mean	Y Geo Mean
UL	1	0.01	0.01	9.73	9.73	16.25	16.25
UR	88	0.89	0.88	409.27	295.75	32.02	26.82
LL	4755	48.06	47.55	5.55	4.78	2.12	1.80
LR	5050	51.04	50.50	111.88	82.55	2.28	1.75

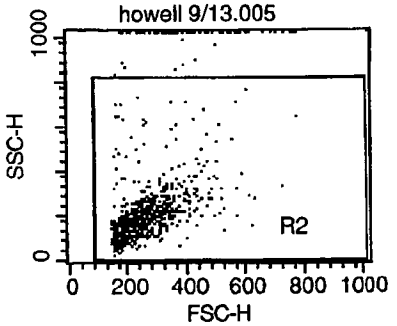


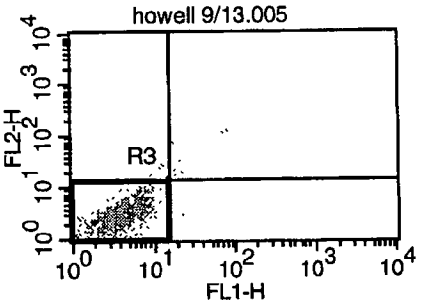
Histogram Statistics

File: howell 9/13.004
Sample ID:
Tube:
Acquisition Date: 13-Sep-99
Gated Events: 9894
X Parameter: FL1-H (Log)

Log Data Units: Linear Values
Patient ID:
Panel:
Gate: G2
Total Events: 10000

Marker	Left, Right	Events	% Gated	% Total	Mean	Geo Mean	CV	Median	Peak Ch
All	1, 9910	9894	100.00	98.94	63.41	21.23	147.72	19.99	4
M1	17, 9910	5065	51.19	50.65	118.42	86.38	88.29	82.05	57



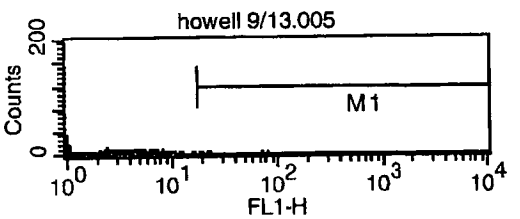


Quadrant Statistics

File: howell 9/13.005
Sample ID: #1 sort
Tube:
Acquisition Date: 13-Sep-99
Gated Events: 689
X Parameter: FL1-H (Log)
Quad Location: 16, 14

Log Data Units: Linear Values
Patient ID:
Panel:
Gate: G2
Total Events: 765
Y Parameter: FL2-H (Log)

Quad	Events	% Gated	% Total	X Mean	X Geo Mean	Y Mean	Y Geo Mean
UL	4	0.58	0.52	12.06	11.86	21.07	20.44
UR	7	1.02	0.92	34.70	28.24	61.07	44.34
LL	676	98.11	88.37	4.28	3.73	3.23	2.77
LR	2	0.29	0.26	19.06	18.85	2.78	2.78

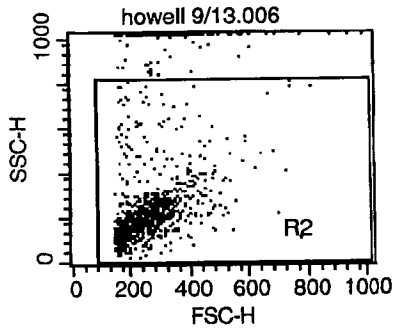


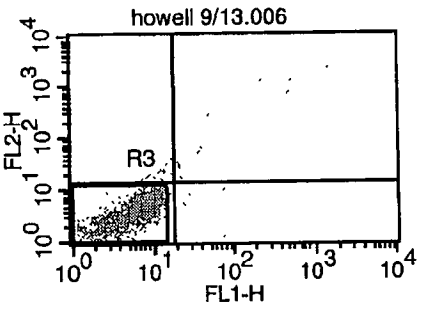
Histogram Statistics

File: howell 9/13.005
Sample ID: #1 sort
Tube:
Acquisition Date: 13-Sep-99
Gated Events: 689
X Parameter: FL1-H (Log)

Log Data Units: Linear Values
Patient ID:
Panel:
Gate: G2
Total Events: 765

Marker	Left, Right	Events	% Gated	% Total	Mean	Geo Mean	CV	Median	Peak Ch
All	1, 9910	689	100.00	90.07	4.68	3.85	97.80	3.89	1
M1	17, 9910	8	1.16	1.05	33.10	27.35	74.74	21.38	17



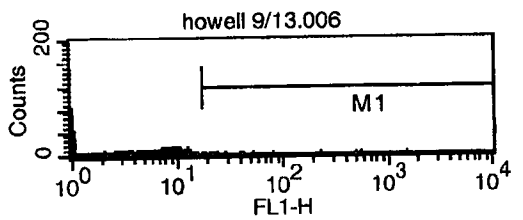


Quadrant Statistics

File: howell 9/13.006
Sample ID: #2 sort
Tube:
Acquisition Date: 13-Sep-99
Gated Events: 1000
X Parameter: FL1-H (Log)
Quad Location: 18, 14

Log Data Units: Linear Values
Patient ID:
Panel:
Gate: G2
Total Events: 1155
Y Parameter: FL2-H (Log)

Quad	Events	% Gated	% Total	X Mean	X Geo Mean	Y Mean	Y Geo Mean
UL	14	1.40	1.21	12.71	12.27	22.33	21.52
UR	11	1.10	0.95	257.57	81.91	566.15	157.95
LL	971	97.10	84.07	5.93	4.69	4.01	3.28
LR	4	0.40	0.35	50.43	43.62	6.32	4.88



Histogram Statistics

File: howell 9/13.006
Sample ID: #2 sort
Tube:
Acquisition Date: 13-Sep-99
Gated Events: 1000
X Parameter: FL1-H (Log)

Log Data Units: Linear Values
Patient ID:
Panel:
Gate: G2
Total Events: 1155

Marker	Left, Right	Events	% Gated	% Total	Mean	Geo Mean	CV	Median	Peak Ch
All	1, 9910	1000	100.00	86.58	8.98	4.95	561.56	5.78	1
M1	17, 9910	18	1.80	1.56	171.58	55.20	201.93	31.91	17