

## V79 COLONY FORMING ASSAY

Experiment Name :  $^3\text{H}_2\text{O}$  + 10% DMSO; Exp. # : 3; Investigator: A. Bishayee  
 Date: 07/13/98

1. Set the rocker-roller at 37°C incubator, set the Coulter Counter, wash cells (from 75 cm<sup>2</sup> flusk, subcultured 1:2, 24h before) with PBS, trypsinize cells, resuspend in 7 ml MEMB, pass five times through 3 cc syringe with 21 gauge needle, perform cell count by transferring 100 ul in Coulter cup containing 20 ml isotone (Coulter balanced electrolyte solution)
2. Dilute to ~4,00,000 cells/ml in MEMB (final volume 11 ml) [Actual count : 379733 cells/ml]
3. Transfer 1 ml of cell suspension into ten 12 ml tubes (Falcon plastic test tube, 17x100 mm) labeled 1-10 both on cap and wall
4. Roll the tubes for 3-4 h at 37°C, 5% CO<sub>2</sub> Date/Time: 07/13/98; 4-00 P.m.
5. Obtain  $^3\text{H}_2\text{O}$  from refrigerator (25 mCi/ml) NEN Catalog # NET001C
6. After 3-4 h, remove test tubes from roller and add MEMB and/or  $^3\text{H}_2\text{O}$  according to Table below. Date/Time: 07/13/98; 7-45 P.m.

Tube #	$^3\text{H}_2\text{O}$ mCi/ml	Cells in MEMB (ml)	MEMB (ul)	$^3\text{H}_2\text{O}$ (ul) [25 mCi/ml]	DMSO (ul)	MEMB (ul)	
1	0	1.0	800	0	200	0	
2	0	1.0	800	0	200	0	
3	0.25	1.0	780	20	200	0	
4	0.75	1.0	740	60	200	0	
5	1.25	1.0	700	100	200	0	
6	0	1.0	800	0	0	200	
7	0	1.0	800	0	0	200	
8	0.25	1.0	780	20	0	200	
9	0.75	1.0	740	60	0	200	
10	1.25	1.0	700	100	0	200	

7. Return test tubes to roller for 12 h, increase the elevation angle of the roller.

Date/Time: 07/13/98; 8-00 P.m.

8. While test tubes are in roller, obtain sterile DMSO (100%) from refrigerator, thaw it, move roller to 10.5°C, obtain ice
9. After ~12 h incubation period, remove tubes from incubator, chill on ice
10. Add DMSO (while vortexing) or MEMB according to the Table, vortex, quickly return to ice  
Date/Time : 07/14/98; 9-00 a.m.
11. Transfer tubes to roller at 10.5 °C for 72 h. Date/Time: 07/14/98; 9-10 a.m.
12. After 72 h, remove tubes, place on ice and centrifuge at 2000 rpm at 4°C for 10 min  
(precooled centrifuge) Date/Time: 07/17/98; 1-30 a.m.
13. Transfer 10 ul medium to test tubes containing 490 ul MEMB in each
14. Add 8 ml ice-cold wash MEMA, vortex
15. Centrifuge tubes for 10 min at 2000 rpm, 4°C
16. Labeling and preparation of dilution tubes and colony dishes
  - load 48 mm petri dishes with 4 ml MEMA
  - load 30 T-tubes with 4.5 ml MEMA and label them 1.2, 1.3, 1.4, 2.2, 2.3, 2.4, X.2, X.3, X.4, etc.
17. Decant supernatant, click tubes, vortex, resuspend in 10 ml wash MEMA
18. Centrifuge tubes for 10 min at 2000 rpm, 4°C
19. Decant supernatant, click tubes, vortex, resuspend in 10 ml wash MEMA
20. Centrifuge tubes for 10 min at 2000 rpm, 4°C
21. Decant supernatant, click tubes, vortex, resuspend in 10 ml wash MEMA
22. Centrifuge tubes for 10 min at 2000 rpm, 4°C
23. Decant supernatant, click tubes, vortex, resuspend in 2 ml wash MEMA, pass five times through 3 cc syringe with 21 gauge needle
24. Determine cell concentration by transferring 100 µl to Coulter cup
25. Vortex tube, transfer 0.5 ml into dilution tube X.4, vortex tube X.4 and transfer 0.5 ml to tube X.3, vortex tube X.3 and transfer 0.5 ml to tube X.2 and vortex. Keep tubes on ice.
26. Transfer 1 ml from dilution tubes into dishes labeled X.2, X.3, X.4 (in triplicate). Only X.2 should be seeded for control T-tubes.
27. Transfer 100 µl of cell suspension (in triplicate) to prelabelled vial (C) for each tube  
containing 3 ml liquid scintillation cocktail  
and count them
28. Incubate petridishes for 1 week
29. ~~Add 490 ul MEMB in tubes containing 10 ul of medium (step 13)~~, vortex, transfer 10 ul in triplicate into prelabelled vials (M).  
containing 3 ml liquid scintillation cocktail
30. ~~Add 3 ml liquid scintillation cocktail to vials and count for radioactivity~~
31. After 1 week, wash colonies 3 times with normal (1X) saline, and 2 times with methanol. Stain colonies with crystal violet
32. Count colonies (50 or more cells). There must be between 25 and 250 colonies for the dish to be a valid data point.

Expt #3

07/13/98

$$\begin{aligned} \text{Initial Cell count} &= 5327, 5416, 5208 \\ \text{Avg. cell count} &= 5317 \\ \text{Cell conc.} &= 5317 \times 400 \\ &= 2,126,800 \text{ cells/ml} \end{aligned}$$

For dilution,

$$\begin{aligned} \text{Vol. of cell suspension required} &= \frac{4400000}{2126800} \\ &= 2.06 \text{ ml} \end{aligned}$$

Take 2.06 ml of cell suspension + 8.94 MEMB = 11 ml.

After dilution,

$$\begin{aligned} \text{Final cell count} &= 955, 976, 917 \\ \text{Avg. cell count} &= 949.3 \\ \text{Cell conc.} &= 949.3 \times 400 \\ &= 379,733 \text{ cells/ml} \end{aligned}$$

USER:10 ID:TRITIUM      PRESET TIME: 1.00      FRI 17 JUL 1998 16:36  
 SAMPLE REPEAT: 1 CYCLE REPEAT: 1 SCR:N      RS232:N  
 : 1 AQC:N QCF:N RCM:N 2 PHASE MONITOR:N  
 CHANNEL 1-LL: 0 UL: 400 2SIGMA: 2.00 BKG SUB: 0.00 BKG 2SIG: 0.00 LSR: 0  
 DATA CALC: CPM, UNKNOWN REPLICATES: 1      NORM FACTOR: 0.100000  
 HALF LIFE(DAYS):N

SAM	POS	CH	CPM	2SIG%	TIME	EL TIME	AVG H#	ERR
1	**	1	21.00	43.64	1.00	1.67	54.0	
2	**	2	25.00	40.00	1.00	3.42	55.0	
3	**	3	43.00	30.50	1.00	5.20	56.0	
4	**	4	27.00	38.49	1.00	6.93	57.0	
5	**	5	37.00	34.82	1.00	8.72	57.0	
6	**	6	33.00	33.81	1.00	10.49	58.0	
7	**	7	33.00	33.33	1.00	12.22	59.0	
8	**	8	45.00	30.50	1.00	14.06	60.0	
9	**	9	60154.29	1.95	0.17	15.01	59.0	
10	**	10	39803.33	1.83	0.30	16.03	56.0	
11	**	11	63870.00	1.77	0.20	16.96	58.0	
12	**	12	172893.33	1.24	0.15	17.84	57.0	
13	**	13	190140.00	1.18	0.15	18.73	58.0	
14	**	14	186560.00	1.20	0.15	19.63	58.0	
15	**	15	303611.44	0.87	0.17	20.60	58.0	
16	**	16	204125.72	0.87	0.17	21.58	59.0	
17	**	17	1725.72	0.92	0.17	22.56	58.0	
18	**	18	43.00	30.50	1.00	24.30	62.0	
19	**	1	27.00	38.49	1.00	26.14	61.0	
20	**	2	32.00	35.36	1.00	27.87	63.0	
21	**	3	36.00	33.33	1.00	29.66	62.0	
22	**	4	30.00	36.51	1.00	31.39	61.0	
23	**	5	46.00	29.49	1.00	33.12	57.0	
24	**	6	40730.91	1.89	0.28	34.17	61.0	
25	**	7	54257.78	.81	0.23	35.17	61.0	
26	**	8	46024.00	.86	0.25	36.14	61.0	
27	**	9	177805.72	.13	0.17	37.11	63.0	
28	**	10	174646.00	1.24	0.15	37.98	61.0	
29	**	11	17529.00	1.14	0.17	38.95	60.0	
30	**	12	30183.00	0.94	0.15	39.85	60.0	
31	**	13	3027.00	0.37	0.17	40.83	58.0	
32	**	14	30545.56	0.93	0.15	41.74	62.0	
37	**	1	34.00	34.30	1.00	43.61	84.0	
38	**	2	45.00	29.81	1.00	45.39	80.0	
39	**	3	42.00	30.86	1.00	47.17	86.0	
40	**	4	24.00	40.82	1.00	48.91	85.0	
41	**	5	29.00	37.14	1.00	50.64	85.0	
42	**	6	46.00	29.49	1.00	52.37	85.0	
43	**	7	1553.00	5.08	1.00	54.16	85.0	
44	**	8	1561.00	5.06	1.00	55.94	86.0	
45	**	9	1533.00	5.11	1.00	57.67	86.0	
46	**	10	3509.00	3.38	1.00	59.46	84.0	
47	**	11	3487.00	3.39	1.00	61.19	84.0	
48	**	12	3714.00	3.28	1.00	62.97	85.0	
49	**	13	6317.00	2.52	1.00	64.72	87.0	
50	**	14	6445.00	2.49	1.00	66.45	86.0	

Handwritten notes and brackets in the table, including 'back', '1M', '2M', '13M', '14M', '15M', '16M', '17M', '18M', '19M', '20M', '21M', '22M', '23M', '24M', '25M', '26M', '27M', '28M', '29M', '30M', '31M', '32M', '1c', '2c', '3c', '4c', and '5c'.

SAM	POS	CH	CPM	2SIG%	TIME	EL TIME	AVG H#	ERR
51	**	-15	6664.00	2.45	1.00	68.18	85.0	
52	**	-16	33.00	34.82	1.00	69.93	83.0	
53	**	-17	35.00	33.81	1.00	71.67	80.0	
54	**	-18	28.00	37.80	1.00	73.45	73.0	
55	**	-1	32.00	35.36	1.00	75.27	86.0	
56	**	-2	38.00	32.44	1.00	77.03	86.0	
57	**	-3	41.00	31.23	1.00	78.82	86.0	
58	**	-4	1546.00	5.09	1.00	80.55	86.0	
59	**	-5	1616.00	4.98	1.00	82.29	84.0	
60	**	-6	1446.00	5.26	1.00	84.03	81.0	
61	**	-7	4245.00	3.07	1.00	85.82	86.0	
62	**	-8	4060.00	3.14	1.00	87.60	80.0	
63	**	-9	4167.00	3.10	1.00	89.33	84.0	
64	**	-10	7050.00	2.38	1.00	91.08	82.0	
65	**	-11	6969.00	2.40	1.00	92.87	79.0	
66	**	-12	7213.00	2.35	1.00	94.61	83.0	

TABLE-1

Expt. # : 3

Date/Time : 07/17/98; 4-36 p.m.

Tube #	Medium count for 10 ul of (cpm) <i>1:50 diluted medium</i>	Avg. cpm	dpm [cpm/0.52]	$\mu$ Ci/ml (A.) on counting [dpm/444]	$\mu$ Ci/ml (A.) <del>on addition</del> <del>[dpm/444]</del>
1	20, 4, 10	11.3	21.7	0.04	0.00004
2	12, 13, 20	15	28.8	0.06	0.00006
3	60131, 39780, 63847	54586	104973	236.4	0.236
4	172870, 190117, 186537	183174.6	352258	793.3	0.793
5	303588, 304102, 268702	292130	561789.7	1265.2	1.265
6	20, 4, 9	11	21.1	0.04	0.00004
7	13, 7, 23	14.3	27.5	0.06	0.00006
8	40707, 54234, 46001	46980	90347	203.4	0.203
9	177782, 174623, 175274	175893	338255	761.8	0.761
10	301810, 302771, 305443	303341	583348	1313.8	1.313

200 = 10

TABLE-2

Expt. # : 3

Date/Time : 07/17/98; 4-36 p.m

Tube #	Radioactivity for 100 ul cell suspension (cpm)	Avg. cpm	dpm [cpm/0.52]	$\mu$ Ci/ml (A <sub>1</sub> ) on counting [dpm/222000]	$\mu$ Ci/ml (A <sub>0</sub> ) after 12 h incubation [A <sub>1</sub> e <sup>-<math>\lambda</math>t</sup> ]
1	11, 22, 19				
2	1, 6, 23				
3	1530, 1538, 1510	1526	2934.6	0.0132	
4	3486, 3464, 3691	3547	6821.1	0.0307	
5	6294, 6422, 6641	6452.3	12408.3	0.0558	
6	10, 12, 5				
7	9, 15, 18				
8	1523, 1593, 1423	1513	2909.6	0.0131	
9	4222, 4037, 4144	4134.3	<del>475</del> 7950.5	0.0358	
10	7027, 6946, 7190	7054.3	13566	0.0611	

**TABLE-3**

Expt. # : 3

Date/Time : 07/17/98; 3-30 p.m

Tube #	Coulter count for 100 ul cell suspension	Avg. count	Cells/ml [Avg. count x 400]	pCi/cell [uCi/ml x 10 <sup>6</sup> Cells/ml]
1	<del>707</del> , 636, 607, 620	621	248400	
2	669, 675, 659	667	267066	
3	712, 680, <del>633</del> , 722	704	281866	0.0468
4	960, 1040, 1021	1007	402800	0.0762
5	1170, 1243, 1077	1163	465333	0.1199
6	1263, 1191, 1202	1218	487466	
7	861, 842, <del>722</del> , <del>767</del> , 806	836	334533	
8	1219, 1249, <del>1348</del> 1269	1245.6	498240	0.0262
9	<del>1110</del> , <sup>830</sup> <del>1013</del> , <sup>880</sup> <del>1001</del> , 866	875.3	350133	0.1022
10	1279, 1266, 1220	1255	502000	0.1217



TABLE-4

Expt # : 3

Date : 07/24/98

Tube.dilution	Colony 1	Colony 2	Colony 3	Avg Colony for x.2	SF
1.2	161	169	153	} 156.33	
2.2	150	149	156		
3.2	117	120	113	116.66	0.7462
4.2	78	70	82	76.66	0.4904
5.2	40	45	36	40.33	0.2579
6.2	140	149	152	} 144	
7.2	139	141	143		
8.2	57	50	64	57	0.3958
9.3	180	188	196	18.8	0.1305
10.4	244	240	248	2.44	0.0169

Expt. #3

3H<sub>2</sub>O + 10% DMSO

DMF = 3:1

10  
12-183  
Made in U.S.A.

SF

0.1

0.01

0.001

Semi-Logarithmic  
3 Cycles x 10 to 1/10 inch

